Lung Tumors in Strain A Mice: Application for Studies in Cancer Chemoprevention

Gary D. Stoner, PhD¹, Gabriela Adam-Rodwell, PhD², and Mark A. Morse, PhD¹

¹ The Ohio State University, Department of Preventive Medicine, Arthur G. James Cancer Hospital and Research Institute, Columbus, OH 43210

² Medical College of Ohio, Department of Pathology, Toledo, OH 43699

Abstract Strain A mice develop a high incidence of spontaneous lung tumors during their lifetime. These tumors may be found in some animals as early as 3 to 4 weeks of age, increasing to nearly 100% by 24 months of age. The strain A mouse is also highly susceptible to the induction of lung tumors by several classes of chemical carcinogens and has been used extensively as a mouse lung tumor bioassay for assessing the carcinogenic activity of a variety of chemicals.

In addition to its use in carcinogen detection, the strain A mouse lung tumor model has been employed extensively for the identification of inhibitors of chemical carcinogenesis. A number of chemopreventive agents including β -naphthoflavone, butylated hydroxyanisole, ellagic acid, phenethyl isothiocyanate, phenylpropyl isothiocyanate, phenylbutyl isothiocyanate, phenylhexyl isothiocyanate, indole-3-carbinol, *etc.*, have been shown to inhibit chemically induced lung tumors in strain A mice. In most instances, inhibition of lung tumorigenesis has been correlated with effects of the chemopreventive agent on the metabolic activation and/or detoxification of carcinogens. To date, no chemopreventive agent has been shown to inhibit lung tumorigenesis in strain A mice when administered after the carcinogen, *i.e.*, during the promotion/progression stages of tumor development. Efforts should be made to develop a standardized protocol in strain A mice for evaluating chemopreventive agents as inhibitors of both the initiation and progression stages of lung tumor development. © 1993 Wiley-Liss, Inc.

Key words: Mouse, lung tumors, isothiocyanates, 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone, benzo(a)pyrene, chemoprevention, NNK, strain A, ras

Strain A mice develop a high incidence of spontaneous lung tumors during their lifetime. These tumors may be found in some animals as early as 3 to 4 weeks of age with a steady increase to nearly 100% by 24 months of age [1]. The strain A mouse is also highly susceptible to the induction of lung tumors by several classes of chemical carcinogens compared to other inbred mouse strains, and has been used extensively as a mouse lung tumor bioassay to assess the carcinogenic activity of a variety of chemicals [2,3].

The histomorphology [1,2,4] and ultrastructure [5] of spontaneous pulmonary neoplasms in strain A mice have been described. However, in these preliminary papers, much emphasis was placed upon the morphological characteristics of the benign tumor (adenoma), with minor reference to the histomorphologic features of the malignant counterpart. In recent articles, Dixon *et al.* [6] described the morphological features of malignant spontaneous neoplasms of strain A mouse lung and classified these as either papillary carcinoma or mixed carcinoma. Foley *et al.* [7] found that chemically induced tumors in

Address reprint requests to Gary D. Stoner, PhD, The Ohio State University, Department of Preventive Medicine, Arthur G. James Cancer Hospital and Research Institute, Room 1148, 300 West 10th Avenue, Columbus, Ohio 43210.

strain A mouse lung arise in hyperplasias and progress to adenoma, carcinoma within adenoma, and ultimately, to carcinomas. Although a matter of some controversy, it would appear that most lung tumors in mice arise from type II cells of the alveolar epithelium [8], with a minor component of papillary tumors arising either from Clara cells of the terminal bronchioles or highly undifferentiated type II cells [9].

The biochemical, growth and transplantation characteristics of mouse lung tumors were summarized in detail by Shimkin and Stoner [1]. A recent study has shown that strain A lung tumors stain intensely for lactate dehydrogenase, NADPH diaphorase, and glyceraldehyde-3-phosphate dehydrogenase activities [10]. Phosphatidylglycerol, an important component of pulmonary surfactant, is present in lower quantities in lung tumors than in normal lung tissue [11]. The proliferative rate of type II alveolar epithelial cells in untreated and chemically treated strain A mice is significantly higher than in mouse strains resistant to lung tumorigenesis [12]. Lung tumor susceptibility in strain A mice also appears to be influenced by the H-2 major histocompatibility complex [13].

The development of spontaneous and chemically induced lung tumors in strain A mice is associated with mutational activation of the Kras oncogene [14, 15]. This appears to be an early event in lung tumorigenesis. In spontaneous tumors, mutations in codons 12, 13 or 61 of the gene appear to occur randomly, whereas carcinogens induce mutations predominately in specific nucleotides of these codons [14,16]. Interestingly, lung tumor susceptibility in strain A mice is correlated with a 37 base pair deletion in the second intron of the K-ras gene [17]. The mechanism(s) by which this deletion influences lung tumor susceptibility is unclear. Other genetic alterations in strain A mouse lung tumors include reductions in the expression of the retinoblastoma (Rb) gene and the growth arrest-specific (gas-3) gene [18]. Preliminary results suggest that the p53 suppressor gene is mutated only in some carcinomas and not in adenomas of strain A mouse lung (Ming You, personal communication).

Lung tumors in mice have been used extensively for the detection of inhibitors of chemical carcinogenesis. A number of chemopreventive agents including β -naphthoflavone [19,20], butylated hydroxyanisole [21], ethoxyquin [22], quercetin [19], pentamethylether [19], sodium cyanate [23], benzyl isothiocyanate [24], phenethyl isothiocyanate [25,26], phenylpropyl isothiocyanate [25,26], phenylbutyl isothiocyanate [25,26], phenylhexyl isothiocyanate [26], ellagic acid [27], tannic acid [28], indole-3-carbinol [29], D-limonene [30], sulindac [31], green tea and black tea [32], and biochanin A [33] have been shown to inhibit chemically induced lung tumors in mice (Table I). Most of these studies have employed strain A mice, although other mouse strains with high to intermediate susceptibility to lung tumorigenesis have also been used. Several protocols have been employed to evaluate the ability of putative chemopreventive agents to inhibit lung tumorigenesis in strain A mice; however, a thorough discussion of these is beyond the scope of this paper. Instead, we present a protocol used in our laboratories, and in the laboratories of our collaborators, Stephen S. Hecht and Fung-Lung Chung at the American Health Foundation, Valhalla, New York, to test chemopreventive agents in the strain A mouse lung tumor model.

MATERIALS AND METHODS

Animals. Four to six week-old male and female A/J mice are purchased from Jackson Laboratories, Bar Harbor, Maine. Male or female mice can be used in chemoprevention protocols since the lung tumor response in A/J mice to carcinogens is similar in both sexes [1]. The mice are maintained in quarantine on a regular laboratory chow diet for two weeks before use in the bioassays. All mice are housed in Bioclean[®] laminar flow rooms in groups of four in solid bottom and side polycarbonate cages. Tap water is available ad libitum. The mice are maintained under standard conditions $(20 \pm 2^{\circ}C; 50 \pm 10\% \text{ relative humidity}; 12)$ hour cycle of light and darkness). Hygienic conditions are maintained by twice-weekly changes of the cages and water bottles, and the cages are sanitized routinely.

<u>Chemicals</u>. All chemicals tested for inhibitory activity in the lung tumor bioassay are stored as recommended by the manufacturer. Carcinogens are usually obtained from the National Cancer Institute's Chemical Carcinogen Repository at the Midwest Research Institute, Kansas City,

Carcinogen	Inhibitor	Reference
Benzo(<i>a</i>)pyrene	β -naphthoflavone	[19]
Benzo(a)pyrene	BHAª, ethoxyquin	[21]
7,12-DMBAª	ВНА	[21]
7-Hydroxymethyl-12- methylbenz (a) anthracene	ВНА	[21]
$\mathrm{Dibenz}(a,h)$ anthracene	ВНА	[21]
Diethylnitrosamine	BHA, ethoxyquin	[21]
4-NQO ^a	BHA, ethoxyquin	[21]
Uracil mustard	ВНА	[21]
Urethane	ВНА	[21]
3-Methylcholanthrene	eta-naphthoflavone	[20]
Benzo(a)pyrene	sodium cyanate	[23]
Benzo(a)pyrene	ellagic acid	[27]
Benzo(a)pyrene	benzyl isothiocyanate	[24]
NNKª	phenethyl isothiocyanate, 3-phenylpropyl isothiocyanate, 4-phenylbutyl isothiocyanate	[25]
Benzo(a)pyrene	tannic acid	[28]
NNK ^a	indole-3-carbinol	[29]
NNK ^a	phenethyl isothiocyanate, 3-phenylpropyl isothiocyanate, 4-phenylbutyl isothiocyanate, 5-phenylpentyl isothiocyanate, 6-phenylhexyl isothiocyanate	[26]
NNK ^a	D-limonene, orange oil, lemon oil	[30]
NNK	sulindac	[31]
NNK	green tea, black tea	[32]
Benzo(<i>a</i>)pyrene	biochanin A	[33]

TABLE I. Inhibitors of Carcinogen-Induced Lung Tumors in Mice

 a 7,12-DMBA = 7,12-dimethylbenz(a)anthracene; BHA = butylated hydroxyanisole; 4-NQO = 4-nitroquinoline-1-oxide; NNK = 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone.

MO, and stored at 4°C in the dark. Chemopreventive agents and carcinogens are tested for purity by high-performance liquid chromatography (HPLC).

<u>Diet</u>. AIN-76A modified diet containing 20% casein, 0.3% DL-methionine, 52% cornstarch, 13% dextrose, 5% corn oil, 5% Alphacel, 3.5% AIN mineral mixture, 1% AIN vitamin mixture,

and 0.2% choline bitartrate is used for routine bioassays of chemopreventive agents in the A/J mouse lung tumor model. Fresh batches of diet are obtained at monthly intervals, and the diet is stored routinely at 4° C in the dark.

Administration of Chemopreventive Agents. Test compounds are administered ig (by gavage). A major advantage of the gavage route is that it allows accurate quantitation of dose. However, repeated administration of test compounds by gavage is labor intensive and can lead to significant mortality in treated animals. An alternative method was suggested in the studies of Castonguay *et al.* [34] who administered the test compound in the diet and the carcinogen in the drinking water.

Bioassay. Groups of 6 to 7 week-old A/J male or female mice are administered the test compound in corn oil by gavage at 3-5 concentrations for four consecutive days. These concentrations are chosen from preliminary dose range-finding studies based upon the observation that they elicit no clinical or other (e.g., weight loss) signs of toxicity. The carcinogen is dissolved in saline (if soluble) or corn oil and administered in a single intraperitoneal (ip) dose within 2-4 hours after the last dose of test compound. Controls consist of mice treated with four consecutive ig doses of the test compound only at the above concentrations, or a single ip dose of carcinogen only. Vehicle controls consist of mice given four consecutive ig doses of corn oil plus one ip dose of either saline or corn oil. The mice are housed four per cage as described above. AIN-76A diet and tap water are provided ad libitum.

The mice are observed daily during the first week of the study, when the treatments are administered, and weekly thereafter. Body weights are measured weekly for the first month and monthly thereafter. All mice are sacrificed and necropsied 16 weeks after administration of the tobacco-specific nitrosamine, 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK) or 24 weeks after treatment with benzo-(a) pyrene [B(a)P]. Sacrifice of B(a)P-treated animals at the later time point (i.e., 24 weeks)appears to be necessary to permit the tumors to grow large enough to be visible on the lung surface. The lungs are infused with saline and fixed in Tellyesniczky's fixative [1]. Surface tumors are counted following fixation, using a dissecting microscope. Representative tumor samples are embedded in paraffin and stained with hematoxylin and eosin for histopathological examination.

Statistical comparisons of tumor multiplicities among the various groups are performed by analysis of variance (ANOVA) followed by Newman-Keuls' ranges test. Comparisons of the proportions of animals in groups that develop tumors are performed by the Chi-square test. Body weight and weight gain are analyzed using one-way analysis of variance followed by Tukey's test when significant differences are observed.

<u>DNA Adduct Analysis</u>. Investigations with NNK have shown a linear relationship between NNK-induced lung tumorigenesis in A/J mice and the formation of O^6 -methylguanine (O^6 -MeGua) adducts in lung DNA [35]. Therefore, studies have been conducted in our laboratories to determine the relationship between inhibition by chemopreventive agents (isothiocyanates and indole-3-carbinol) of NNK-induced lung tumors in A/J mice and reduction in the formation of O^6 -MeGua in lung DNA.

Groups of A/J mice are administered corn oil or chemopreventive agents by gavage for four consecutive days. On the fourth day, NNK is administered ip 2 hours after the final gavage. All treatments are identical to those used in the bioassay. Groups of 5 animals are sacrificed at 2 hours or 6 hours following NNK dosing. DNA is isolated from excised lungs by a modification of the method of Marmur [36] and purified by the method of Sebti et al. [37]. The DNA is hydrolyzed in 0.1N HCl for 30 minutes at 70°C to release O⁶-MeGua and guanine. 7-MeGua, O⁶-MeGua, and guanine are quantitated by strong cation exchange HPLC coupled with fluorescence detection. The identities of 7-MeGua, O⁶-MeGua, and guanine are confirmed by coelution with authentic standards.

RESULTS AND DISCUSSION

Examples of data obtained from bioassays of chemopreventive agents in the A/J mouse lung tumor model are shown in Tables II, III, and IV. In Table II, a series of arylalkyl isothiocyanate compounds of increasing chain length were evaluated by Morse et al. [25,26] for their ability to inhibit lung tumorigenesis by the tobaccospecific carcinogen, NNK. Female A/J mice were administered either phenyl isothiocyanate (PITC), benzyl isothiocyanate (BITC), phenethyl isothiocyanate (PEITC), 3-phenylpropyl isothiocyanate (PPITC), 4-phenylbutyl isothiocyanate (PBITC), 5-phenylpentyl isothiocyanate (PPeITC), or 6-phenylhexyl isothiocyanate

Treatment ¹	No. of mice	Tumors/mouse ^{c,d}	% of Mice with tumors ^e
Corn oil + NNK	39	9.2 ± 0.5^{1}	100 ¹
PITC + NNK	30	9.8 ± 0.9^{1}	100 ¹
BITC + NNK	29	10.4 ± 0.7^{1}	100^{1}
PEITC + NNK	28	3.3 ± 0.4^2	93^{1}
PPITC + NNK	30	0.4 ± 0.1^{3}	37^2
PBITC + NNK	28	0.4 ± 0.1^3	32^{2}
Corn oil + NNK	60	7.9 ± 0.4^{1}	100 ¹
PEITC + NNK	20	4.1 ± 0.8^2	93^{2}
PPITC + NNK	19	0.2 ± 0.1^4	11^{3}
PBITC + NNK	19	0.2 ± 0.1^4	11^{3}
PPeITC + NNK	20	0.3 ± 0.1^4	25^3
PHITC + NNK	20	0.1 ± 0.1^4	5^3

TABLE II. Effects of Isothiocyanates on NNK-induced Lung Tumors in Strain A/J Mice^a

^aData of Morse *et al.* [43]: Groups of 20–40 strain A/J mice were administered corn oil or isothiocyanates (5 μ mol/mouse/day) by gavage daily for four consecutive days. Two hours after the final dose of corn oil or test compounds, a single dose of saline or NNK (10 μ mol/mouse) in saline was administered ip. Sixteen weeks after NNK administration, mice were killed and pulmonary tumors counted.

^bPITC = phenyl isothiocyanate; BITC = benzyl isothiocyanate; PEITC = phenethyl isothiocyanate; PPITC = phenylpropyl isothiocyanate; PBITC = phenylbutyl isothiocyanate; PPITC = phenylpentyl isothiocyanate; PHITC = phenylhexyl isothiocyanate.

 $^{\circ}$ Mean \pm S.E.

^dMeans in this column that bear different superscripts are significantly different (p < 0.05) from one another.

 $^{\circ}$ Percentages in this column that bear different superscripts are significantly different (p < 0.0001) from one another.

(PHITC) by gavage for 4 consecutive days at doses of 5, 1, and 0.2 μ mol/day prior to administration of 10 μ mol of NNK by ip injection. Mice were sacrificed 16 weeks after NNK administration and lung tumors were quantitated. When compared to vehicle (corn oil + saline) controls, none of the isothiocyanate compounds were shown to influence the occurrence of spontaneous lung tumors in A/J mice (Table II). Animals treated with NNK only had an average of 9.2 \pm 0.5 (mean \pm S.E.) lung tumors. PITC and BITC were ineffective as inhibitors of NNK lung tumorigenesis. PEITC effectively inhibited NNK-induced lung tumors at a dose of 5 μ mol/day but

was not inhibitory at doses of 1 or 0.2 μ mol/day. PPITC, PBITC, PPeITC, and PHITC were all more potent inhibitors of NNK lung tumorigenesis than PEITC. PHITC appeared to be the most potent tumor inhibitor of all of the isothiocyanates. At a dose of 0.2 μ mol/day, PHITC treatment reduced the tumor multiplicity by 85% (data not shown). These results support the hypothesis that increased alkyl chain length enhances the inhibitory activity of an arylalkyl isothiocyanate towards NNK lung tumorigenesis.

DNA adduct studies provided data in support of the lung tumor results. Groups of A/J mice

Treatment	O ⁸ -MeGua (µmol/mol guanine) ^b		
Corn oil	17.6 ± 0.9^{1}		
PEITC	17.2 ± 0.2^{1}		
PPITC	13.8 ± 0.4^2		
PBITC	13.4 ± 0.5^2		
PPeITC	11.1 ± 0.2^3		
PHITC	9.9 ± 0.2^{3}		

TABLE III. Effects of Isothiocyanates on NNK-induced DNA Methylation in Lungs of Strain A/J Mice^a

^aData of Morse *et al.* [26]. Groups of 15 strain A/J mice were administered corn oil or isothiocyanates (1 μ mol in 0.1 ml corn oil) by gavage for four consecutive days. At 2 hours after the final treatment, mice were given 10 μ mol NNK ip. Mice were killed 6 hours after NNK administration and their lungs excised. Following DNA isolation and purification, O⁶MeGua was analyzed as described in **MATERIALS AND METHODS**. Values within the same column that bear different superscripts are statistically different from one another (p < 0.05).

^bMean \pm S.E.

Treatment	Dose (µmol)	No. of mice	Tumors/mouse ^b	% of Mice with tumors			
Corn oil + tricaprylin		48	0.6 ± 0.9	40			
PEITC	1.5	46	0.6 ± 0.9	40			
PEITC	15.0	47	0.4 ± 0.7	32			
$B(a)P$ (8 μ mol)		41	8.4 ± 6.9	100			
PEITC + $B(a)P$	1.5	42	10.5 ± 9.2	100			
PEITC + $B(a)P$	5.0	46	12.3 ± 12.0	97			
PEITC + $B(a)P$	10.0	42	9.4 ± 6.0	100			
PEITC + $B(a)P$	15.0	40	7.7 ± 6.7	91			

 TABLE IV. Effects of Phenethyl Isothiocyanate (PEITC) on B(a)P-induced Lung

 Tumors in Strain A/J Mice^a

^aGroups of 24 male and 24 female A/J mice were administered corn oil or phenethyl isothiocyanate (1.5, 5, 10, and 15 μ mol/mouse/day in corn oil) by gavage daily for six consecutive days. At 2 hours after the fourth treatment, mice were given 8 μ mol B(a)P in tricaprylin ip. Twenty four weeks after B(a)P administration, mice were killed and pulmonary tumors counted.

^bMean \pm S.D.

were administered corn oil vehicle or isothiocyanates $(1 \ \mu mol in \ 0.1 \ ml corn oil)$ by gavage for 4 consecutive days. At 2 hours after the final pretreatment mice were administered 10 µmol NNK (in 0.1 ml saline) ip. The animals were sacrificed 6 hours after NNK administration and the levels of O⁶-MeGua in lung DNA were determined as described in MATERIALS AND METHODS. O⁶-MeGua formation was not significantly affected by PEITC at a dose of $1 \mu mol/day$ (Table III). In contrast, both PPITC and PBITC significantly reduced formation of O⁶-MeGua in lung DNA below that of NNKtreated controls and PEITC-pretreated mice. PPeITC and PHITC both significantly reduced O^6 -MeGua levels below those of PPITC and PBITC. Therefore, the relative potency for inhibiting NNK-induced lung DNA methylation was found to be: PHITC \approx PPeITC > PBITC \approx PPITC > PEITC. This order is in substantial agreement with that observed for inhibitors of NNK-induced lung tumorigenesis.

An explanation for the differences in the abilities of arylalkyl isothiocyanates to inhibit NNK-induced lung tumorigenesis has been discussed [26]. PEITC has been shown to inhibit the cytochrome P-450-dependent microsomal metabolism of nitrosamines, including NNK [25,38]. When added to lung microsomes in vitro, the inhibitory activity of arylalkyl isothiocyanates towards NNK α -hydroxylation was found to increase with increasing chain length [38]. With increasing alkyl chain length for a given isothiocyanate, lipophilicity increases and reactivity decreases [26], both of which may affect the delivery of the compound to the lung. In addition, increased chain length may favor binding of an isothiocyanate to the catalytic site(s) of cytochrome P-450 isozyme(s) responsible for NNK α -hydroxylation. At present, at least two cytochrome P-450s, *i.e.*, P-450IA1 and P-450IIB, appear to be involved in NNK metabolism in mouse lung [38].

Table IV illustrates data obtained when PEITC was evaluated for its ability to inhibit B(a)P tumorigenesis in A/J mouse lung. Groups of mice were administered PEITC by gavage for 6 consecutive days at doses of 1.5, 5, 10, and 15 μ mol/day. A single dose of 8 μ mol of B(a)P in 0.1 ml tricaprylin was given by ip injection after the fourth dose of PEITC. Mice were sacrificed 24 weeks after B(a)P administration and lung tumors were quantitated. When compared to vehicle (corn oil + tricaprylin) controls, PEITC did not influence the occurrence of spontaneous lung tumors in A/J mice. Tumor incidence and multiplicity in the PEITC + B(a)P groups were not significantly different from the B(a)P control groups. Therefore, when tested in a protocol similar to that used for the NNK experiments, PEITC was found to be ineffective as an inhibitor of B(a)P tumorigenesis in A/J mouse lung.

The basis for the lack of inhibitory effect of PEITC towards B(a)P-induced lung tumors in A/J mice is unknown. It is possible that differences in the metabolism of NNK and B(a)P may account for the observed differences between PEITC activity toward NNK- and B(a)P-induced lung tumors. In addition, the treatment regimen and dose levels selected for PEITC and B(a)P may have masked the inhibitory potential of PEITC against B(a)P-induced lung tumors. Pharmacokinetic studies have shown that PEITC and NNK are rapidly absorbed and eliminated in mice [29,39]. In contrast, B(a)Padministered ip is eliminated slowly from mice [40]. Therefore, the persistence of B(a)P in the animals for long periods after the elimination of the inhibitor may have been responsible for the lack of PEITC activity. Wattenberg [41] demonstrated inhibitory effects of PEITC against DMBA-induced lung and forestomach tumors in mice. In this study [41], the inhibitor and carcinogen were mixed in the diet and administered to mice for a period of four weeks. This treatment regimen may be more appropriate when using carcinogens that are slowly eliminated from animals, and in situations in which it is desirable to maintain a favorable ratio of the concentration of inhibitor to that of the carcinogen.

To date, no chemopreventive agent has been shown to inhibit lung tumorigenesis in strain A mice when administered after the carcinogen, *i.e.*, during the promotion/progression stages of tumor development. Agents found to be effective as inhibitors of lung tumorigenesis have generally been shown to influence the metabolic activation and/or detoxification of carcinogens, *i.e.*, events associated with tumor initiation. In our opinion, efforts should be made to develop a standardized protocol to evaluate chemopreventive agents as inhibitors of both the initiation and progression stages of lung tumor development in strain A mice.

Historically, the strain A mouse lung tumor model has been criticized because of its perceived lack of "relevance" to human lung cancer [1,2,3]. Most tumors arising in the terminal airways of the mouse lung have been classified as benign adenomas, as opposed to the invasive, malignant carcinomas seen in human lung. However, the aforementioned studies of Dixon et al. [6] and Foley et al. [7] clearly indicate that both spontaneous and chemically induced lung tumors in strain A mice progress from adenoma \rightarrow carcinoma within adenoma \rightarrow carcinoma with increasing age of the animals. The histopathological features of mouse lung carcinomas resemble those of adenocarcinomas arising in the terminal airway of human lung. Moreover, mouse lung tumors and at least 40% of human lung adenocarcinomas have at least one molecular marker in common, i.e., activation of the Kras oncogene [14,16]. Recent epidemiological studies have revealed an increase in the frequency of human lung adenocarcinomas [42]; therefore, the strain A mouse lung tumor model may assume more importance as a relevant model for a type of human lung cancer that is becoming more prevalent.

REFERENCES

- Shimkin MB, Stoner GD: Lung tumors in mice: Application to carcinogenesis bioassay. Adv Cancer Res 21:1–58, 1975.
- Stoner GD, Shimkin MB: Strain A mouse lung tumor bioassay. J Am Coll Toxicol 1:145-169, 1982.
- Stoner GD: Lung tumors in strain A mice as a bioassay for carcinogenicity of environmental chemicals. Exp Lung Res 17:405-423, 1991.
- Grady HG, Stewart HL: Histogenesis of induced pulmonary tumors in strain A mice. Am J Pathol 16:417-437, 1940.
- Brooks RE: Pulmonary adenoma in strain A mice: An electron microscopic study. J Natl Cancer Inst 41:719-742, 1968.
- Dixon D, Horton J, Haseman JK, Talley F, Greenwell A, Nettesheim P, Hook GE, Maronpot RR: Histomorphology and ultrastructure of spontaneous pulmonary neoplasms in strain A mice. Exp Lung Res 17:131-155, 1991.
- Foley JF, Anderson MW, Stoner GD, Gaul BW, Hardisty JF, Maronpot RR: Proliferative lesions in the mouse lung: Progression studies in strain A mice. Exp Lung Res 17:157-168, 1991.
- 8. Rehm S, Devor DE, Henneman JR, Ward JM: Ori-

gin of spontaneous and transplacentally induced mouse lung tumors from alveolar type II cells. Exp Lung Res 17:181–195, 1991.

- Thaete LG, Malkinson AM: Cells of origin of primary pulmonary neoplasms in mice: Morphologic and histochemical studies. Exp Lung Res 17:219-228, 1991.
- Gunning WT, Stoner GD, Goldblatt PJ: Glyceraldehyde-3-phosphate dehydrogenase and other enzymatic activity in normal mouse lung and in lung tumors. Exp Lung Res 17:255-261, 1991.
- 11. Stoner GD, Hallman MN, Troxell MC: Lecithin biosynthesis in a clonal line of lung adenoma cells with type II alveolar cell properties. Exp Mol Pathol 29:102-119, 1978.
- Thaete LG, Beer DG, Malkinson AM: Genetic variation in the proliferation of murine pulmonary type II cells: Basal rates and alterations following urethane treatment. Cancer Res 46:5335-5338, 1986.
- Oomen LCJM, van der Valk MA, Demant P: MHC and non-MHC genes in lung tumor susceptibility in the mouse: Implications for the study of the different lung tumor types and their cell of origin. Exp Lung Res 17:283-304, 1991.
- You M, Maronpot RR, Stoner GD, Anderson MW: Activation of the K-ras protooncogene in chemically induced lung tumors of the strain A mouse. Proc Natl Acad Sci 86:3070-3074, 1989.
- Belinsky SA, Devereux TR, Maronpot RR, Stoner GD, Anderson MW: The relationship between the formation of promutagenic adducts and the activation of the K-ras protooncogene in lung tumors from A/J mice treated with nitrosamines. Cancer Res 49:5305-5311, 1989.
- You M, Wang Y, Lineen A, Stoner GD, You L, Maronpot RR, Anderson MW: Activation of protooncogenes in mouse lung tumors. Exp Lung Res 17:389-400, 1991.
- You M, Wang Y, Maronpot RR, Stoner GD, Anderson MW: Partial bias of K-ras oncogenes detected in lung tumors from mouse hybrids. Proc Natl Acad Sci 89:5804-5808, 1992.
- Re FC, Manenti G, Borrello MG, Colombo MP, Fisher JH, Pierotti MA, Porta GD, Dragani TA: Multiple molecular alterations in mouse lung tumors. Mol Carcinog 5:155-160, 1992.
- Wattenberg LS, Leong JL: Inhibition of the carcinogenic action of benzo(a)pyrene by flavones. Cancer Res 30:1922–1925, 1970.
- Anderson LM, Priest LJ: Reduction in the transplacental carcinogenic effect of methylcholanthrene in mice prior to treatment with β-naphthoflavone. Res Commun Chem Pathol Pharmacol 30:431-446, 1980.
- Wattenberg LW: Inhibition of chemical carcinogeninduced pulmonary neoplasia by butylated hydroxyanisole. J Natl Cancer Inst 50:1541-1544, 1973.
- 22. Wattenberg LW: Inhibition of carcinogenic and toxic effects of polycyclic hydrocarbons by phenolic anti-

oxidants and ethoxyquin. J Natl Cancer Inst 48: 1425-1430, 1972.

- Wattenberg LW: Inhibition of polycyclic aromatic hydrocarbon-induced neoplasia by sodium cyanate. Cancer Res 40:232-234, 1980.
- Wattenberg LW: Inhibitory effects of benzyl isothiocyanate administered shortly before diethylnitrosamine or benzo(a)pyrene on pulmonary and forestomach neoplasia in A/J mice. Carcinogenesis 8: 1971-1973, 1987.
- Morse MA, Amin SG, Hecht SS, Chung F-L: Effects of aromatic isothiocyanates on tumorigenicity, O⁶methylguanine formation, and metabolism of the tobacco-specific nitrosamine 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone in A/J mouse lung. Cancer Res 49:2894-2897, 1989.
- Morse MA, Eklind KI, Hecht SS, Jordan KG, Choi C-I, Desai DH, Amin SG, Chung F-L: Structureactivity relationships for inhibition of 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK) lung tumorigenesis by arylalkyl isothiocyanates in A/J mice. Cancer Res 51:1846-1850, 1991.
- 27. Lesca P: Protective effects of ellagic acid and other plant phenols on benzo(a)pyrene-induced neoplasia in mice. Carcinogenesis 4:1651–1653, 1983.
- Athar M, Khan WA, Mukhtar H: Effect of dietary tannic acid on epidermal, lung and forestomach polycyclic aromatic hydrocarbon metabolism and tumorigenicity in Sencar mice. Cancer Res 49: 5784-5788, 1989.
- 29. Morse MA, LaGreca SD, Amin SG, Chung F-L: Effects of indole-3-carbinol on lung tumorigenesis and DNA methylation induced by 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK) and on the metabolism and disposition of NNK in A/J mice. Cancer Res 50:2613-2617, 1990.
- Wattenberg LW, Coccia JB: Inhibition of 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone carcinogenesis in mice by D-limonene and citrus fruit oils. Carcinogenesis 12:115-117, 1991.
- Pepin P, Bouchard L, Nicole P, Castonguay A: Effects of sulindac and oltipraz on the tumorigenicity of 4-(methylnitrosamino)1-(3-pyridyl)-1-butanone in A/J mouse lung. Carcinogenesis 13:341-348, 1992.
- Wang ZY, Hong J-Y, Huang M-T, Reuhl KR, Conney AH, Yang CS: Inhibition of N-nitrosodiethylamine- and 4-(methylnitrosamino)-1-(3-pyri-

dyl)-1-butanone-induced tumorigenesis in A/J mice by green tea and black tea. Cancer Res 52:1943– 1947, 1992.

- Lee YS, Kim TH, Cho KJ, Jang JAJ: Inhibitory effects of biochanin A on benzo(a)pyrene induced carcinogenesis in mice. In Vivo 6:283-286, 1992.
- Castonguay A, Pepin P, Stoner GD: Lung tumorigenicity of NNK given orally to A/J mice: Its application to chemopreventive efficacy studies. Exp Lung Res 17:485-499, 1991.
- Peterson LA, Hecht SS: O⁶-methylguanine is a critical determinant of 4-(methylnitrosamino)-1-(3pyridyl)-1-butanone tumorigenesis in A/J mice lung. Cancer Res 51:5557-5564, 1991.
- Marmur J: A procedure for the isolation of deoxyribonucleic acid from microorganisms. J Mol Biol 3:208-218, 1961.
- Sebti SM, Pruess-Schwartz DM, Baird WM: Speciesand length of exposure-dependent differences in the benzo(a)pyrene: DNA adducts formed in embryo cell cultures from mice, rats, and hamsters. Cancer Res 45:1594-1600, 1985.
- Smith TJ, Guo Z, Thomas PE, Chung F-L, Morse MA, Eklind KI, Yang CS: Metabolism of 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone in mouse lung microsomes and its inhibition by isothiocyanates. Cancer Res 50:6817-6822, 1990.
- Eklind KI, Morse MA, Chung F-L: Distribution and metabolism of the natural anticarcinogen phenethyl isothiocyanate in A/J mice. Carcinogenesis 11:2033– 2036, 1990.
- 40. Ross J, Nelson G, Kligerman A, Erexson G, Bryant M, Earley K, Gupta R, Nesnow S: Formation and persistence of novel benzo[a]pyrene adducts in rat lung, liver, and peripheral blood lymphocyte DNA. Cancer Res 50:5088–5094, 1990.
- 41. Wattenberg LW: Inhibition of carcinogenic effects of polycyclic hydrocarbons by benzyl isothiocyanate and related compounds. J Natl Cancer Inst 58:395– 398, 1977.
- 42. Devesa SS, Shaw GL, Blot WJ: Changing patterns of lung cancer incidence by histological type. Cancer Epidemiol Biomarkers Prev 1:29-34, 1991.
- Morse MA, Elkind KI, Amin SG, Hecht SS, Chung FL: Effects of alkyl chain length on the inhibition of NNK-induced lung neoplasia in A/J mice by arylalkyl isothiocyanates. Carcinogenesis 10:1757– 1759, 1989.